

Metabolic characteristics  
in dormant *Mycobacterium  
tuberculosis*——an essential  
physiology shift during latent  
tuberculosis infections.

Ye Yu

Supervised by Prof. GP Zhao

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Department of Microbiology

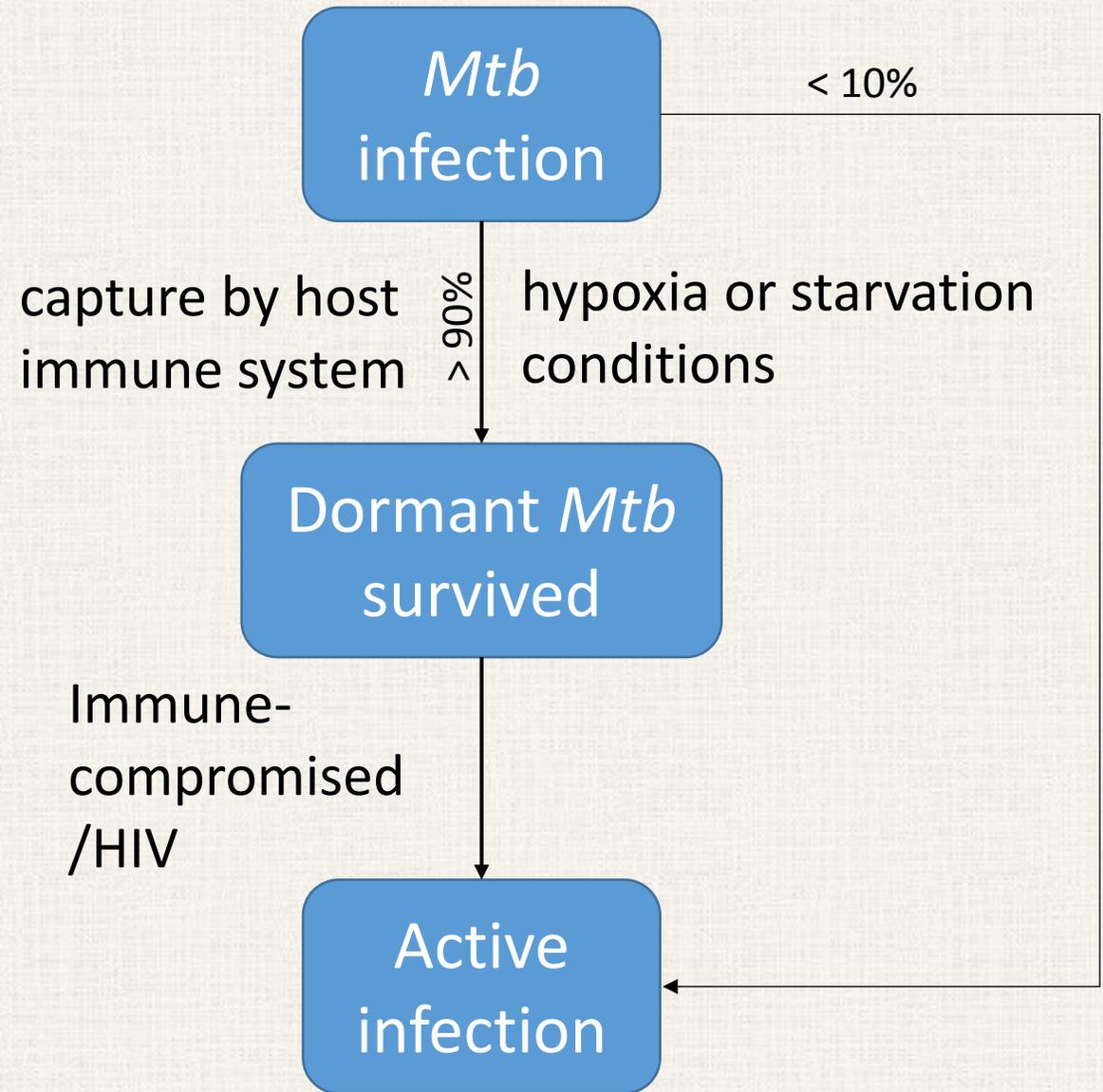
# Outline

- Background of *Mtb* dormancy
  - I. Dormant *Mtb* & LTBI
  - II. Features of Dormant *Mtb*
- Metabolic changes in dormant *Mtb*
  - I. Carbon metabolism
  - II. ATP synthesis
  - III. Respiratory chain
  - IV. Redox balance
- Dormant regulation in *Mtb*
  - I. stringent response
  - II. DosR regulation system
- *Mtb* dormancy & molybdenum cofactor

# Part I. Background of *Mtb* dormancy

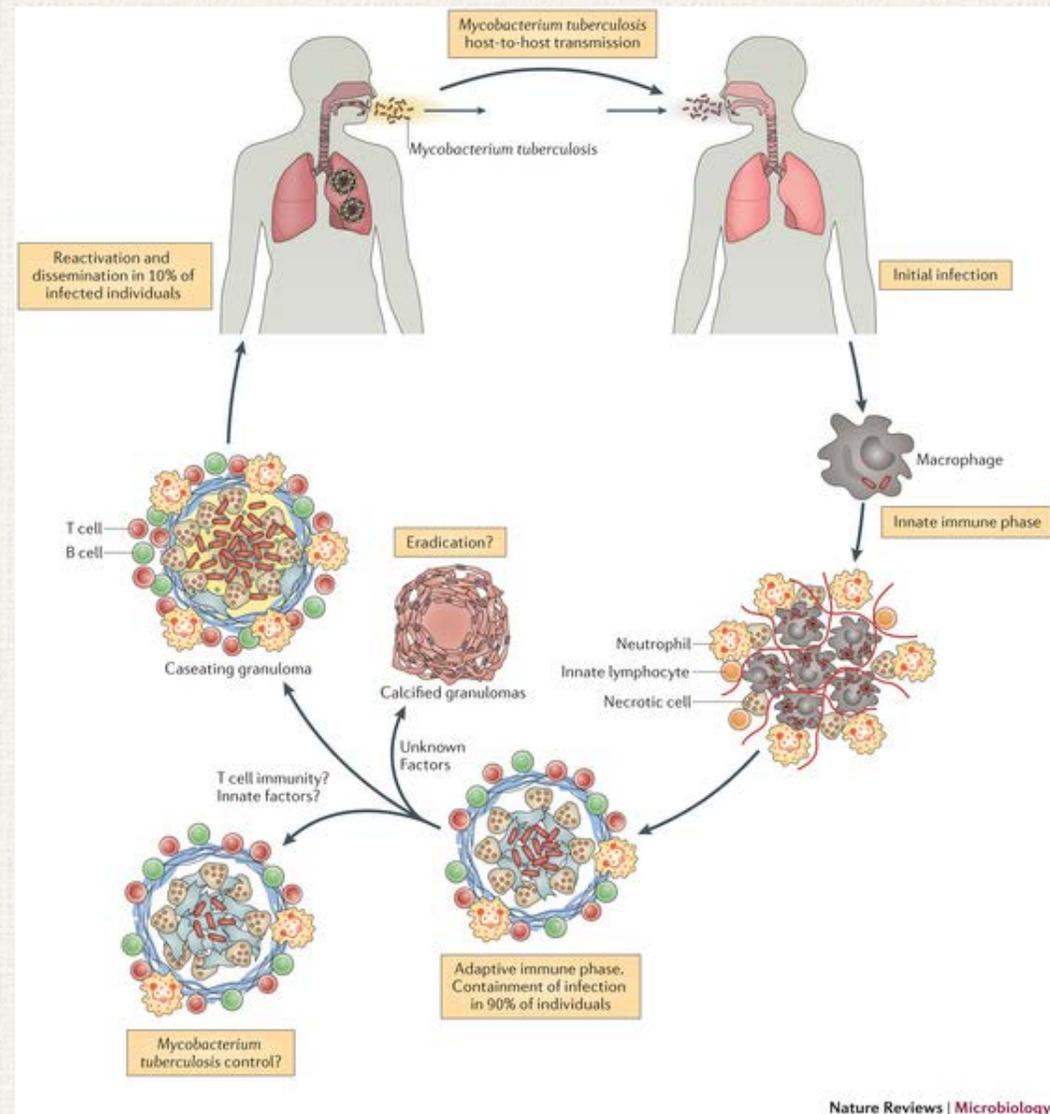
# latent infections of tuberculosis & *Mtb* dormancy

- The incredible **success** of *Mtb* with **Widely spread & long-term treatment** largely due to the **latent infection of tuberculosis (LTBI)**
  - Many proof showed **dormant *Mtb*** is the main state of pathogen during **the latent/persistent infections**
  - Dormant *Mtb* is induced during **infection process** in host cell and might remain latent state for **decades**
- When in more **favorable conditions** → resuscitate to active → active infection



# Advantage of dormancy during *Mtb* infection

- Evade most **immune stress** from host cells **for decades**
- Difficult to cure: at least a **six months treatment & relapse** (remain few dormant cell in entire *Mtb*)
- **Higher** chance to accumulate **Drug-resistance mutants** (multi- or Extensively-drug resistant *tb*)



A typical latent infection of *Mtb*

“captured → multiplication → enter dormancy → living in macrophage → resuscitation → active infection”

# Stress and living environment in host cells

- **Before granuloma formed**

- Immune response (mononuclear cells and T lymphocytes)
- Low pH
- Oxidative stress (NO/CO produced by macrophage)

- **After granuloma matured (solid granuloma)**

- **Hypoxia**
- Low nutrition (foamy macrophage contains rich fatty acid in granuloma center)

# Basic features of dormant *Mtb*

- **Low metabolism level & Non-replicative state**
  - Thickening of the cell wall
  - Shutdown of most transcription & protein synthesis
  - Decrease of **ATP synthesis**
- Maintaining **respiration & the integrity of cell membrane**
  - Switch **electron transfer/ acceptor**
  - Alter **carbon metabolism pathway**
  - Maintain the **proton gradient** across the membrane



Hibernation  
of mammal

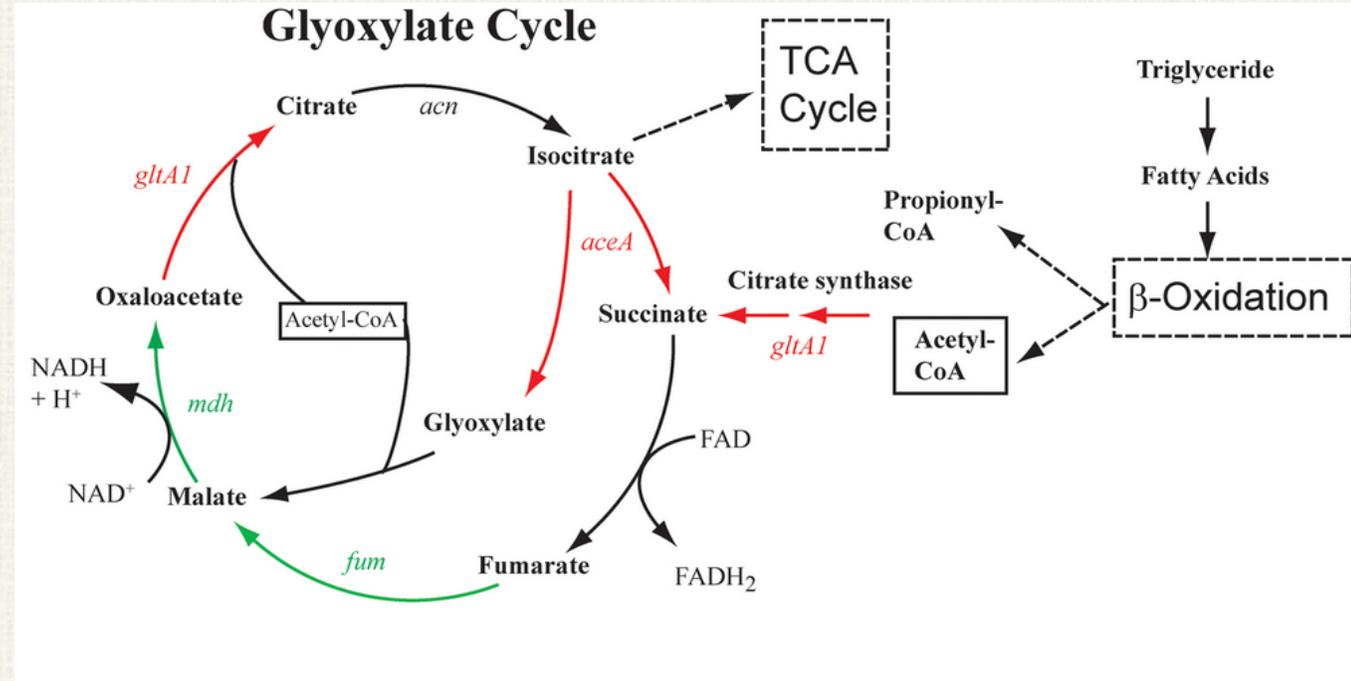


Dormancy of  
microbes

# Part II. Metabolic changes in dormant *Mtb*

# Carbon metabolism

- The dual stresses in macrophage: hypoxia/unbalanced nutrient pool rich in fatty acids and poor in carbohydrates
- **Five-step beta-oxidation pathway & Glyoxylate cycle** was up regulated but **carbohydrates metabolism** was **turned down**
- Isocitrate lyase (ICL) is reported plays another role in suppressing cell apoptosis of macrophage



Microarray analysis demonstrated changes in expression of genes involved in glyoxylate cycle. Daniel J, et al. (2009)

# *ATP synthesis*

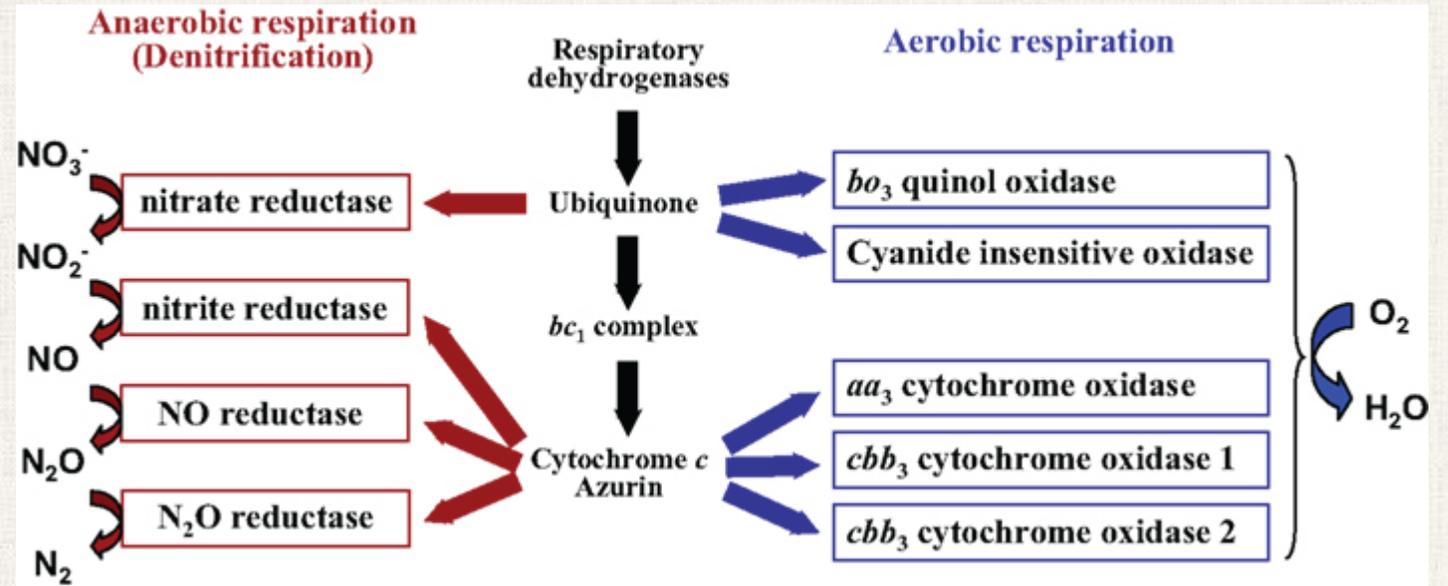
- Cell-wide downregulation of metabolism & nonreplicating
- In the **transcription analysis**, the most significant **ATP synthase cluster atpA-H** strongly down regulated, suggesting a **global down-regulation** of the ATP synthesis
- ATP is a **scarce resource** in the dormant cell

Rv number	Up-regulation Score	Down-regulation Score	Growth-attenuation Score	Gene ID
<b>F1FO ATP synthase</b>				
Rv1308	0	0	-14.68	4.95 atpA
Rv1304	0	0.97	-15.56	4.825 atpB
Rv1311	0	3.465	-9.49	3.66 atpC
Rv1310	0	0	-15.32	4.55 atpD
Rv1305	0	0	-12.87	2.45 atpE
Rv1306	0	0	-15.845	4.745 atpF
Rv1309	0	0	-13.2	4.525 atpG
Rv1307	0	0	-11.865	4.59 atpH

*Scoring and annotations for ATP synthase subunit genes. Searchable table of scoring results for genes encoding various ATP subunits in the Mycobacterium tuberculosis genome. Dennis J Murphy (2007)*

# Change of Respiratory chain

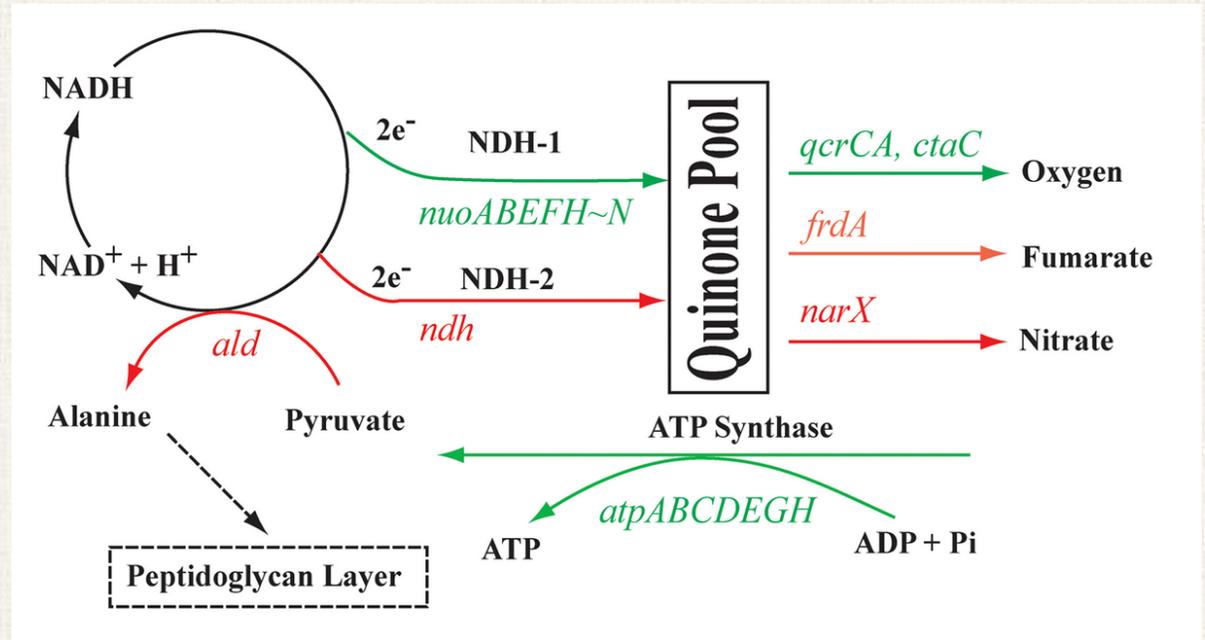
- Lack of terminal electron acceptors (O<sub>2</sub>)
- **Nitrate becomes new main electron acceptors**



- The respiratory chain is also changed, different from **Quinol & cytochrome** transferring the electron in aerobic situation, a series of **nitrogen reductase** form the new anaerobic electron transfer chain
- Nitrate is reduced by a nitrate reductase (*narGHJ*) and is then excreted by a nitrite extrusion protein (*narK1, narK2, narK3*)
- **Alternate** electron carriers in the hypoxic: **fumarate reductase**; probable **NAD(P)H dehydrogenases**; **ferredoxin** (These three parts were upregulated in transcription analysis)

# Redox balance

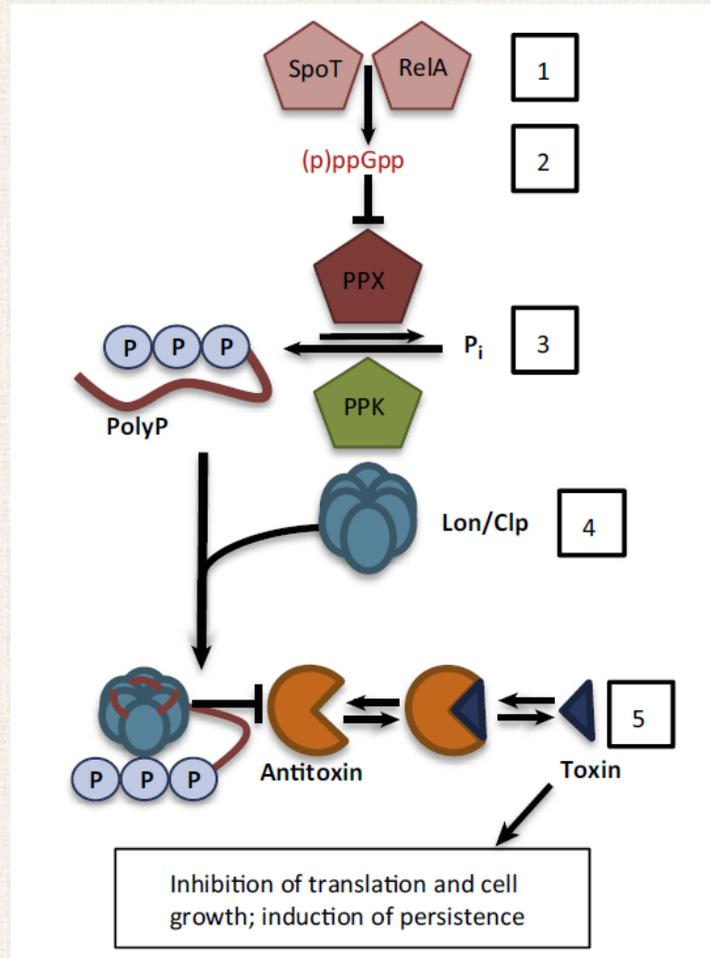
- Lack of terminal **electron acceptors**, NADH dehydrogenase I subunits and ubiquinol–cytochrome C complex were **repressed**
- Beta-oxidation of fatty acid, NADH was produced. To ensure the ratio of NADH/NAD<sup>+</sup>, the NADH dehydrogenase II is **upregulated**. To help the proton transferring, expression of lower-efficiency anaerobic respiration enzymes (*frdA*, *narG/X*, *nirA*) are also **increased**
- Besides, **aminotransferase** is upregulated, expend NADH and take part in other anabolism such as thickening of the cell wall



Microarray analysis demonstrated changes in expression of genes involved in glyoxylate cycle. Daniel J, et al. (2009)

# Part III. Stress sensor & regulation in Mtb dormancy

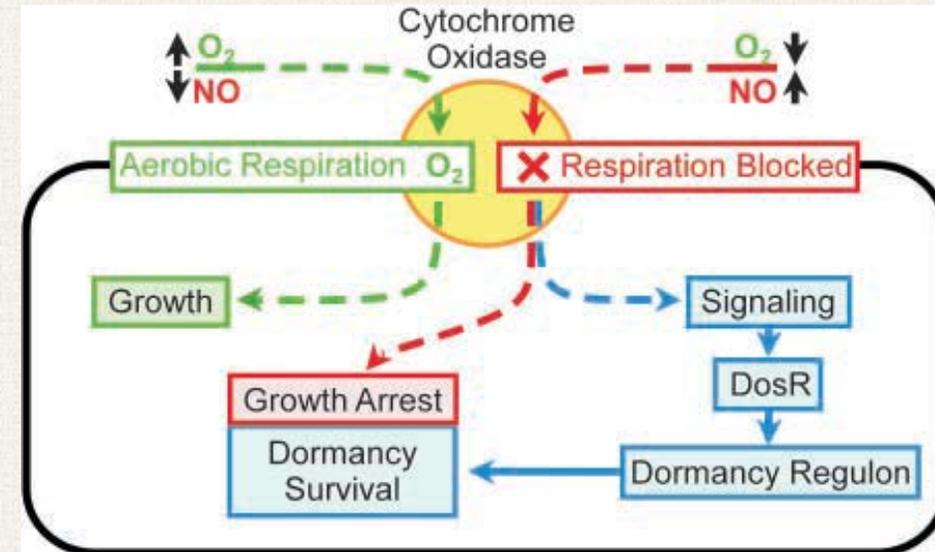
# Stringent response : response to low nutrition



- In *Mtb*, the **ratio** of **amino-acylated tRNA** to **free tRNA** was the first regulatory response to amino acid & carbon starvation by **RelA**
- **ppGpp** is maintained in the cytosol by **RelA**
- ppGpp inhibits **polyphosphatase**, result in the accumulating of **PolyP**. **PolyP** interacts with **TA module**, finally **globally** affect **RNA polymerase**, then down-regulate gene expression

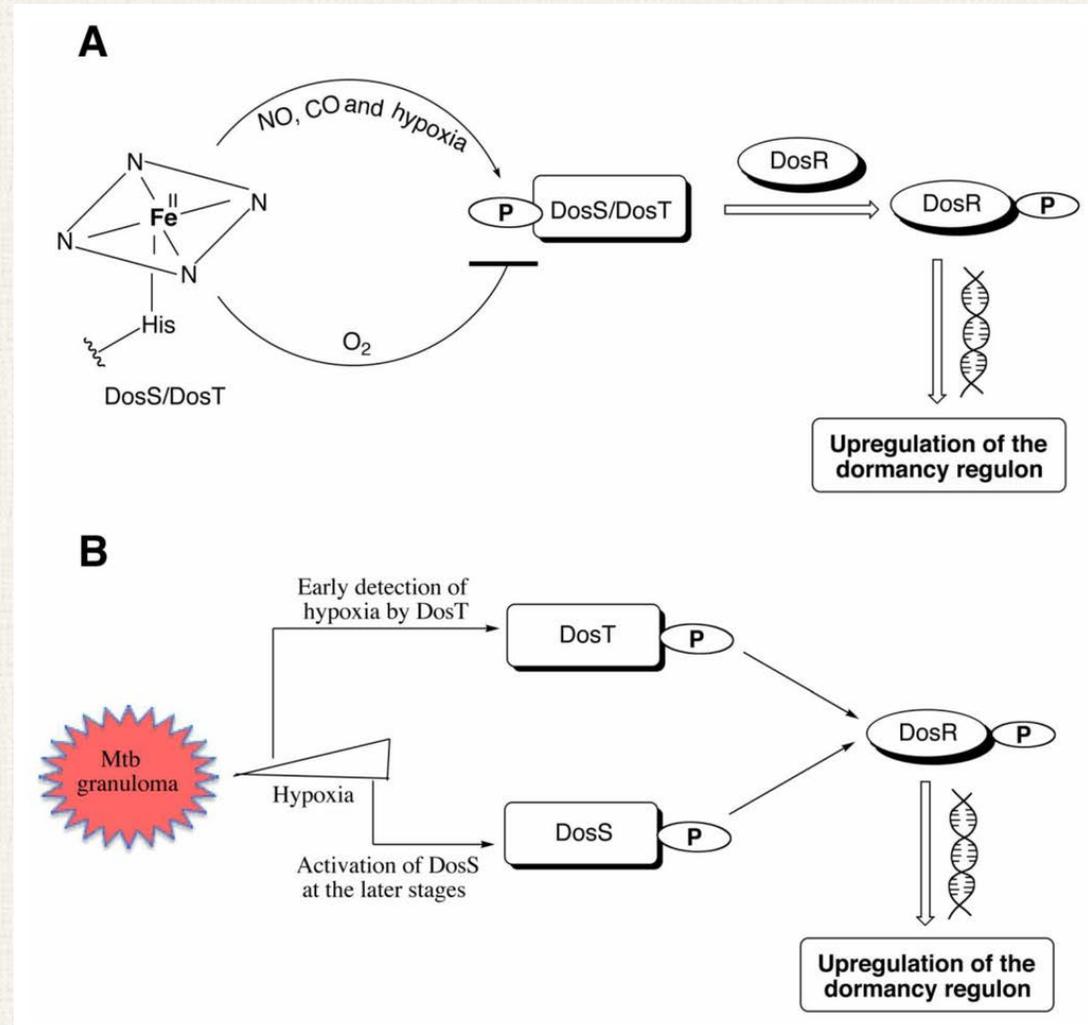
# DosT/DosS two component sensor & DosR regulon : response to hypoxia & Oxidative stress

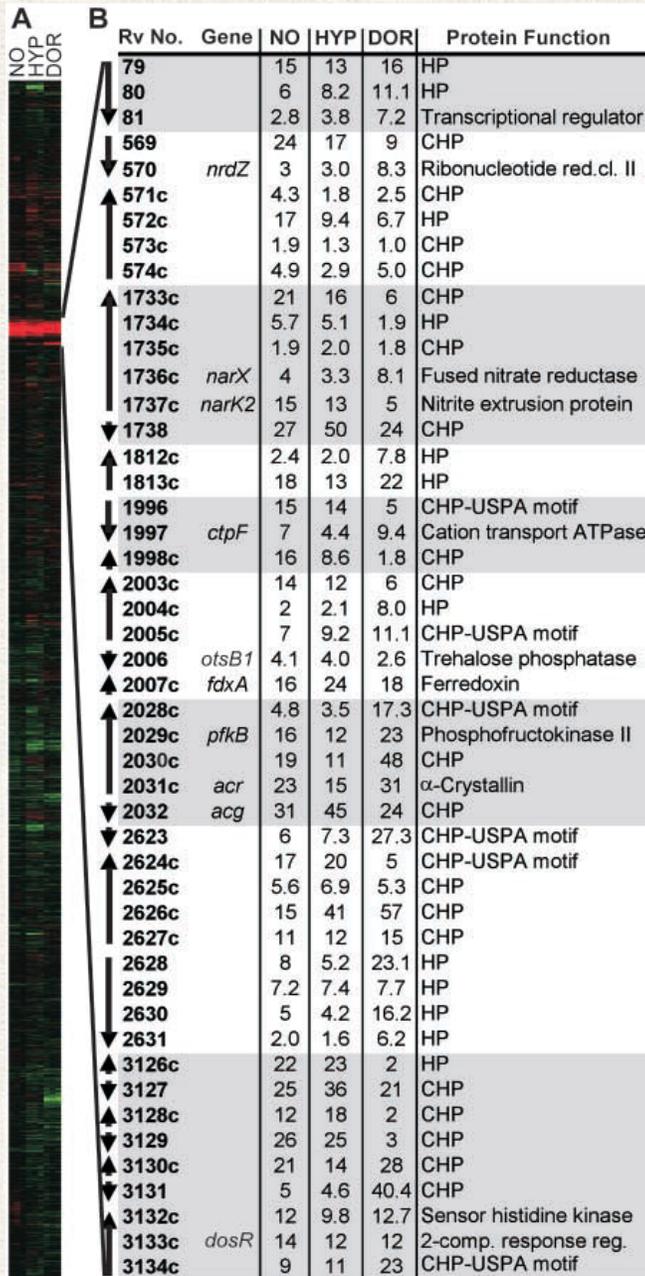
- DosR system seems to be the secondary important dormancy regulator in hypoxia, next to the directly growth arrest by lack of O<sub>2</sub> (red color)
- Expression of DosR was induced by **DosT/DosS** two component sensor
- DosR controls a regulon of **more than 53 genes**, including enzyme in metabolism, respiration pathway, even two regulators



# DosRST two-component system

- DosT is a **gas sensor**, activated by **absence of oxygen** or the binding of **nitric oxide** and **carbon monoxide**. DosS is a **redox state sensor**
- Both DosT/DosS are **Kinase to Phosphorylate** DosR, resulting in downstream signaling
- DosT & DosR activated in different time in hypoxia of granuloma





# DosR regulon

- Now 53 genes was found regulated by *dosR*. Including **4 transporters, 2 Nitrate respiratory chain, 2 regulator** (cascading signal)
- Nearly 60% of the genes do not have an annotated function, by sequence & domain comparison, 11 involved in **carbohydrate and fatty acid metabolism; 8 in electron transfer**

# New insights of the DosR regulon

- In Zheng X' work (2010), DosR showed an additional signaling networks involving **Serine/threonine protein kinases**
- In Thomson NR' work(2011), they discovered **noncoding small RNAs** that appear to be under DosR control.
- In Voskuil MI' work(2013), DosR is required for *M. tuberculosis* **exits the dormant state**

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## Convergence of Ser/Thr and Coordinate Expression of the *Mycobacterium tuberculosis* <sup>3</sup>†

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Joseph D. Chao<sup>1,2</sup>, Kadamba G. Papavinasundaran Guinevere Q. Lee<sup>3</sup>, and Yossef Av-Gay<sup>1,5,4</sup>

From the <sup>1</sup>Department of Microbiology and Immunology and University of British Columbia, Vancouver, British Columbia

OPEN ACCESS Freely available online

## Sequence-Based Analysis Uncovers Non-Coding RNA in the Total T *Mycobacterium tuberculosis*

Kristine B. Arnvig<sup>1\*</sup>, Iñaki Comas<sup>1#</sup>, Nicholas R. Thomson Nicholas J. Croucher<sup>2</sup>, Graham Rose<sup>1</sup>, Timothy T. Perkins<sup>2</sup>, Young<sup>1</sup>

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## The *Mycobacterium tuberculosis* DosR Regulon Assists in Metabolic Homeostasis and Enables Rapid Recovery from Nonrespiring Dormancy<sup>3</sup>†

Rachel L. Leistikow, Russell A. Morton, Iona L. Bartek, Isaac Frimpong, Karleen Wagner, and Martin I. Voskuil\*

University of Colorado Denver, School of Medicine, Department of Microbiology, P18-9115, 12800 East 19th Avenue, P.O. Box 6511, Aurora, Colorado 80045

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# Several Models of Mtb dormancy

- Wayne's anaerobic model
- rather reflects an adaptive response to low oxygen conditions than the state of true deep dormancy
  
- Other models
- based upon culturing *M. tuberculosis* in Sauton's medium without potassium. Under these conditions, more than 99% of bacterial cells transit to dormant, non-culturable state during a prolonged, 60-d stationary phase.
- gradual acidification of the medium, resulting in a massive accumulation of ovoid cells with the properties closely resembling those predicted for dormant bacteria.

# Reference

1. Dennis J Murphy, James R Brown\*. Identification of gene targets against dormant phase *Mycobacterium tuberculosis* infections. **BMC Infectious Diseases** (2007).
2. Calvin Boon & Thomas Dick\*. How *Mycobacterium tuberculosis* goes to sleep: the dormancy survival regulator DosR a decade later. **Future Microbiol** (2012). 7(4), 513–518
3. Martin Gengenbacher & Stefan H.E. Kaufmann. Mycobacterium tuberculosis: success through dormancy. **FEMS Microbiol Rev** 36 (2012) 514–532
4. Shiloh MU, Manzanillo P, Cox JS. *Mycobacterium tuberculosis* senses host-derived carbon monoxide during macrophage infection. **Cell Host Microbe** 3(5), 323–330 (2008).
5. Chao JD, Papavinasasundaram KG, Zheng X *et al.* Convergence of Ser/Thr and two-component signaling to coordinate expression of the dormancy regulon in *Mycobacterium tuberculosis*. **J. Biol. Chem.** 285(38), 29239–29246 (2010).
6. Leistikow RL, Morton RA, Bartek IL, Frimpong I, Wagner K, Voskuil MI. The *Mycobacterium tuberculosis* DosR regulon assists in metabolic homeostasis and enables rapid recovery from nonrespiring dormancy. **J. Bacteriol.** 192(6), 1662–1670 (2010).
7. Watanabe S, Zimmermann M, Goodwin MB, Sauer U, Barry CE 3rd, Boshoff HI. Fumarate reductase activity maintains an energized membrane in anaerobic *Mycobacterium tuberculosis*. **PLoS Pathog.** 7(10), E1002287 (2011).
8. Cole ST, Brosch R, Parkhill J *et al.* Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. **Nature** 393(6685), 537–544 (1998).
9. Dasgupta N, Kapur V, Singh KK *et al.* Characterization of a two-component system, *devR–devS*, of *Mycobacterium tuberculosis*. **Tuber. Lung Dis.** 80(3), 141–159 (2000).
10. Kumar A, Deshane JS, Crossman DK *et al.* Heme oxygenase-1-derived carbon monoxide induces the *Mycobacterium tuberculosis* dormancy regulon. **J. Biol. Chem.** 283(26), 18032–18039 (2008).
11. Commandeur S, Lin MY, Van Meijgaarden KE *et al.* Double- and monofunctional CD4 and CD8 T-cell responses to *Mycobacterium tuberculosis* DosR antigens and peptides in long-term latently infected individuals. **Eur. J. Immunol.** 41(10), 2925–2936 (2011).
12. David R. Sherman and Gary K. Schoolnik *et al.* Inhibition of Respiration by Nitric Oxide Induces a *Mycobacterium tuberculosis* Dormancy Program. **J. Exp. Med.**(2003)
13. Santhosh Sivaramakrishnan and Paul R. Ortiz de Montellano. The DosS-DosT/DosR Mycobacterial Sensor System. **Biosensors** 2013, 3, 259-282; doi:10.3390

*Thank you!*